ORIGINAL ARTICLE

# Particulate matter inhalation during hay storing activity induces systemic inflammation and platelet aggregation

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Abstract The aim of this study was to investigate possible pathomechanisms behind the cardiovascular morbidity caused by inhalation of particulate matter  $(PM_{10})$ . For that purpose, healthy volunteers were exposed to high PM<sub>10</sub> concentrations during a 2 h hay storing activity. Blood was drawn in the evening before and after  $PM_{10}$  exposure and in the morning and evening of the day after exposure. The leukocyte count increased after PM<sub>10</sub> exposure with an initial increase of segmented neutrophils followed by banded forms. C-reactive protein increased over time. Fibrinogen and plasma viscosity became increased in the evening of the day after PM<sub>10</sub> exposure. Platelet aggregation was increased in the evening after  $PM_{10}$  exposure. At the same time von Willebrand factor and factor VIII were increased, reflecting endothelial activation. These results confirm that acute inhalative exposure to high PM<sub>10</sub> concentrations during hay storage activity leads to a systemic inflammatory reaction, endothelial activation, and platelet aggregation.

**Keywords** *C*-reactive protein · Endothelium · Inflammation · Particulate matter · Platelets · Viscosity

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## Introduction

Ambient air pollution is associated with increased cardiovascular mortality (Pope et al. 2004). Such air pollutants are gases or particulate matter (PM), the latter having gained special scientific and public interest in the last few years (Brook et al. 2004). The size of the PM determines the pathophysiological role. PM<sub>10</sub>, also called thoracic particles, have an aerodynamic diameter smaller than 10  $\mu$ m. Upon inhalation they can reach the tracheobronchial tree; wheras larger particles (>10  $\mu$ m) are held off by the nasal epithelium. The smaller the particles are, the deeper they penetrate into the lung periphery, eventually reaching the alveolar space.

Long-term epidemiological studies have consistently shown an association between air pollution and mortality for all causes as well as cardiovascular events (Dockery et al. 1993; Pope et al. 1995). Short-term exposure to high ambient PM<sub>10</sub> pollution leads to higher hospitalisation rates for cardiovascular disease (Hoek et al. 2001), an increased risk for myocardial infarction (Zanobetti and Schwartz 2005), and ischemic stroke (Wellenius et al. 2005). An elevation in PM<sub>10</sub> by 10  $\mu$ g/m<sup>3</sup> was associated with a 4.5% increase in risk for acute ischemic coronary events (Pope et al. 2006), and a 0.2–0.3% increase in daily cardiopulmonary mortality (Dominici et al. 2005).

The pathophysiological mechanisms behind the cardiovascular effects of  $PM_{10}$  are not well understood. Direct effects may occur via agents, which readily cross the pulmonary epithelium into the circulation such as gases, and possibly ultrafine particles along with soluble constituents of  $PM_{2.5}$  (Nemmar et al. 2002). Less acute, indirect effects may occur via a pulmonary oxidative stress with a subsequent systemic inflammatory reaction with increased leucocytes (Salvi et al. 1999), fibrinogen (Ghio et al. 2000) and *C*-reactive protein (Ruckerl et al. 2006), which all may impair vascular functions and accelerate atherosclerosis. Inhalation of organic dust containing toxigenic moulds and mycotoxins during indoor exposure induces an immunological pulmonary reaction (Bünger et al. 2004; Gray et al. 2003).

We have previously shown that farmers in the Swiss Alps are exposed to very high  $PM_{10}$  concentrations during their work, especially hay storage (Cathomas et al. 2002). This gave us the unique opportunity to investigate the influence of a  $\sim 2$  h exposure to such high concentrations of rural  $PM_{10}$  during hay storage in healthy volunteers on parameters of hemorheology, systemic inflammation, plasmatic coagulation and platelet function.

## Methods

# Study design

Fourteen healthy volunteers (six females, eight males, age 26-54 years, median 33.5 years) gave their informed consent to participate in the study, which had been approved by the local Ethics Committee. They were members of the medical staff, non-smokers, in good physical condition and accustomed to the intermediate altitudes of the area. They were exposed in groups of 2-3 to PM inhalation during hay storage with an electrically powered hay blower between 02.00 and 04.00 p.m. on two farms in the Swiss Alps of the Grison: Farm A in Scheid, at 1,300 m above sea level, and Farm B in Cresta, Avers valley, at 1,960 m. The physical activity consisted of loading the hay, which was brought into the barn periodically, from the floor into the blower. Between such periods of work they had large breaks, during which they were relaxing in the barn. The physical stress was more on the muscles than on the cardiovascular system and can be rated as moderate.

# Quantification of PM exposure

To collect the organic dust, an Aerosol Sampler DHA-80 (Digitel, Hegnau, Switzerland) was used, which was positioned in similar distance from the hay blower and level above ground as the faces of the working volunteers were. The filters were weighed before and after exposure at the same conditions (room temperature 20°C). The results were related to the filtered air volume and given as  $\mu g/m^3$ . On the first day at farm B we additionally used a cascade impactor (Anderson Mark II, Particle Size Stack Sampler) to analyse the distribution of the aerodynamic diameters of the particles. The filters were desiccated for 48 h under defined conditions and weighed with an analytical balance (Mettler<sup>®</sup> Toledo, Greifensee, Switzerland).

## Blood sampling

Blood was sampled on four different occasions: at 08.00 p.m. on the day before exposure (Day 0, baseline value), 08.00 p.m. on the day of exposure (Day 1), i.e. 6 h after the end of PM<sub>10</sub> exposure, and in the morning of the day after exposure (Day 2) at 08.00 a.m. and at 08.00 p.m. Each time 5 ml tubes containing 0.106 mol/l buffered sodium citrate, a 4.5 ml tube with lithium–heparin, a 4.5 ml tube for serum and two 4 ml tubes containing EDTA-K (ethylene-diaminetetraacetic acid) were drawn. Plasma and serum samples were deepfrozen at  $-80^{\circ}$ C.

## Hematologic parameters

Erythrocyte and erythrocyte indices, leucocyte and platelet count were measured in blood anticoagulated with EDTA with an electronic particle counter (Sysmex K-1000, Kobe, Japan). Blood smears were prepared, air dried and stained with May-Grünwald-Giemsa.

## Blood and plasma viscosity

Whole blood was incubated at 37.0°C for 10 min and viscosity was measured with a Couette-type viscometer (Contraves LS-30, Mettler-Toledo, Greifensee, Switzerland). We used a high shear rate of 94.5 s<sup>-1</sup> and a low shear rate of 0.1 s<sup>-1</sup> for whole blood, plasma was measured at 11 s<sup>-1</sup>.

# Platelet function analysis

Platelet aggregatory function was tested with a platelet function analyzer (PFA-100<sup>®</sup>, Dade Behring, Düdingen, Switzerland). In this instrument blood is aspirated into a 150  $\mu$ m membrane pore coated with collagen and either epinephrine bitartrate (EPI cartridge) or adenosine diphosphate (ADP cartridge) for platelet activation. Platelets adhere to the pore surface, become activated, then aggregate and finally form an occluding platelet plug in the pore, which stops blood flow (Closure time CT). All measurements were done in duplicate and the mean value was calculated.

### Inflammatory parameters

The plasma concentrations of tumour necrosis factor alpha (TNF-alpha) and interleukin-6 (IL-6) were measured by ELISA (human IL-6, TNF-alpha, R&D Systems, Minneapolis, MN, USA) following the instructions of the manufacturer. *C*-reactive protein (CRP) was measured with a high-sensitivity test (hs-CRP) using latex enhanced nephelometry (BN II, Behring Diagnostics Inc. Marburg, Germany).

# Coagulation parameters

Factor VIII (FVIII) activity was assayed by a standard PTT-based commercial test (Dade Behring, Marburg, Germany) using Prothrombin SL as the contact activating agent. A functional determination of fibrinogen (Multifibren U, Dade Behring, Marburg, Germany) and the ristocetin cofactor activity of von Willebrand factor (BC von Willebrand Reagent, Dade Behring, Marburg Germany) were performed. Plasma thrombin–antithrombin (TAT) complexes were determined using a quantitative enzymoimmunoassay (Enzygnost TAT micro; Dade Behring, Marburg, Germany). Human soluble P selectin (sP-selectin) was measured by a quantitative sandwich solid phase ELISA using a monoclonal antibody directed against soluble P selectin (R&D Systems, Minneapolis, MN, USA).

#### Statistics

A GraphPad prism Version 4.0 for Windows was used for statistical analysis. A one-way analysis of variance (ANOVA) for non-parametric data with Dunn's Multiple Comparison Test for the comparison with baseline values before  $PM_{10}$  exposure were used. A *P*-value < 0.05 was regarded as statistically significant.

## Results

#### PM<sub>10</sub> measurements

The PM<sub>10</sub> concentrations measured near the hay blower during the work of the participants are shown in Table 1. High values were found with a substantial day-to-day variation depending on the weather conditions with low values on days with high humidity and a very high value on the last day, which was after several very warm and dry days. Exposure times varied from 85 to 120 min. The resulting total PM<sub>10</sub> exposure (PM<sub>10</sub> concentration × time) is also shown in Table 1. No significant correlations were found between

773 100% 80% 60% Percentage 40% 20% 0% 10 100 1000 10000 Aerodynamic diameter [nm] - Ambient air (mainly traffic) Ambient air (mainly wood combustion) Hay dust

Fig. 1 Cumulative distribution of the particle aerodynamic diameter of  $PM_{10}$  collected with a cascade impactor during hay storage. For comparison, the particle diameter distributions of ambient air polluted by traffic and wood combustion, measured with the same instrument on other occasions, are shown

the degree of  $PM_{10}$  exposure and any of the parameters below. This may be due to the relatively low number of participants and sometimes large variations, which holds true as well for other parameters mentioned below.

The particle diameter distribution ( $PM_{10}$ -mass) measured in this study near a hay blower is shown in Fig. 1. The PM aerodynamic diameter was between 2 and 10  $\mu$ m, which is about 10 times higher than the diameters measured in air polluted by traffic or wood combustion, as shown in the same figure.

## Hematological data

Erythrocyte count, erythrocyte indices and platelet count were not affected by  $PM_{10}$  exposure (data not shown). The leuco-

**Table 1** Date of investigation, $PM_{10}$  concentrations, exposuretimes, total  $PM_{10}$  exposure andnumber of exposed volunteersworking in two farms with a hayblower on six different days

Date	$PM_{10}  (\mu g/m^3)$	Exposure time (min)	Cumulative PM <sub>10</sub> exposure (mg min/m <sup>3</sup> )	Volunteers ( <i>n</i> )
Farm A				
21/07/2005	10,103	110	1,111	2
21/07/2005	3,960	100	396	3
21/07/2005	7,129	85	606	3
Farm B				
21/07/2005	2,684	107	287	2
21/07/2005	3,695	108	399	2
09/08/2005	14,525	120	1,743	2

08.00 p.m.
$3.77 \pm 1.23$
$0.88\pm0.51$
$2.90\pm0.99$
$0.14\pm0.12$
$0.01\pm0.02$
$0.24\pm0.24$
$2.79\pm0.73$

**Table 2** Differential leucocyte count in absolute numbers before (Day 0) and after  $PM_{10}$  exposure (mean  $\pm$  SD)

\*\* P < 0.01, \*\*\* P < 0.001 compared with baseline on Day 0 (ANOVA)

cyte count was significantly increased in the evening after  $PM_{10}$  exposure due to an increase of polymorphonuclear cells, which is shown in Table 2. An initial rise of segmented neutrophils in the evening after the  $PM_{10}$  exposure was followed by an increase of banded neutrophils in the next morning (Fig. 2). Eosinophils remained unaffected by  $PM_{10}$  exposure.



**Fig. 2** Absolute and differential leucocyte count at 8 p.m. on day 0 (*baseline value*), at 8 p.m. on day 1 after PM10 exposure (*arrow*), and at 8 a.m. and 8 p.m. on day 2. Mean  $\pm$  SD. \* Denotes P < 0.01, \*\*\* P < 0.001 compared with baseline values (ANOVA)

#### Hemorheological data

Hemorheologic data are shown in Table 3. Whole blood viscosity, which is primarily determined by erythrocyte count as well as erythrocyte deformability and aggregation, was not affected by  $PM_{10}$  exposure. Plasma viscosity, which is determined by the concentration of large molecules such as immunoglobulins and fibrinogen, became elevated on day 2 at 8 p.m., and coincided with an increased fibrinogen level at that time (Table 3).

#### Inflammatory markers

Acute  $PM_{10}$  exposure induced a systemic inflammatory response, which is shown by the increase of hs-CRP over time (Fig. 3). The mean value of the whole group (n = 14) was significantly above the baseline value on the post expositional day at 08.00 p.m., i.e. 30 h after  $PM_{10}$  exposure. The large SD on day 2 reflects marked interindividual variations ranging from 0.58 to 38.00 mg/l at 08.00 p.m. These interindividual differences were not related to different  $PM_{10}$  concentrations (Table 1). Systemic serum concentrations of TNF-alpha and IL-6, were not affected by  $PM_{10}$  inhalation (data not shown).

Table 3	Hemorheologic parameters i	n 14 individuals	exposed to high	concentrations of	f PM <sub>10</sub> on day	1
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	Day 0	Day 1	Day 2	
	08.00 pm	08.00 pm	08.00 am	08.00 pm
Whole blood viscosity (mPa s)				
At 94.5 s <sup>-1</sup>	$4.43\pm0.53$	$4.45\pm0.47$	$4.51\pm0.55$	$4.38\pm0.49$
At 0.1 s <sup>-1</sup>	$36.53 \pm 8.22$	$35.51\pm8.10$	$38.01 \pm 9.89$	$35.32\pm8.17$
Plasma viscosity (mPa s)	$1.25\pm0.11$	$1.24\pm0.09$	$1.27\pm0.08$	$1.33\pm0.08*$
Fibrinogen (g/l)	$2.62\pm0.57$	$2.66\pm0.72$	$2.87\pm0.80$	$3.14 \pm 0.90^{**}$

Mean  $\pm$  SD, \* P < 0.05, \*\* P < 0.01 compared with baseline on day 0 (ANOVA)



**Fig. 3** Serum levels of high sensitivity *C*-reactive protein (hs-CRP) before and different time points after  $PM_{10}$  exposure. Mean  $\pm$  SD, n = 14. \*\*\* Denotes P < 0.001 compared with baseline value (ANO-VA)

## Platelet aggregation

The platelet aggregatory function as measured with the PFA-100<sup>®</sup> instrument is shown in Fig. 4. The shorter closure times (CT) with either ADP or epinephrine as a platelet activator indicate increased platelet aggregation after PM<sub>10</sub> exposure. sP-selectin, which is released during platelet aggregation, tended to be higher immediately after PM<sub>10</sub> exposure (23.6 ± 8.0, 28.1 ± 18.0, 25.2 ± 10.1, and 24.2 ± 11.0 ng/ml, respectively).

## Coagulation proteins

The serum levels of vWF and FVIII, which are stored in endothelial cells and released into the circulation upon endothelial cell activation, were increased after  $PM_{10}$  exposure (Fig. 5). The plasmatic coagulation cascade was not initiated by  $PM_{10}$  exposure, as measured by the unchanged levels of thrombin–antithrombin complexes (TAT:  $4.5 \pm 8.4, 2.7 \pm 1.5, 6.2 \pm 12.7, and 2.1 \pm 0.6$  ng/ml at the four different time points of blood sampling, respectively).

# Discussion

Increasing evidence suggests that PM air pollution may adversely affect the cardiovascular system (Brook et al. 2004; Peters 2005). An association between acute  $PM_{10}$  exposure and acute myocardial infarction (Lanki et al. 2006) and hospital admission (Barnett et al. 2006) has been



**Fig. 4** Pore closing time by occluding platelet aggregates using either ADP cartridges (*above*) or EPI cartridges (*below*) on the platelet function analyser PFA-100<sup>®</sup> (see "Methods"). Values (mean  $\pm$  SD, n = 14) are before and various time intervals after PM<sub>10</sub> exposure. Shorter closure times indicate increased platelet aggregation. \* P < 0.05, \*\* P < 0.01, \*\*\* P < 0.001 compared with baseline values (ANOVA)

described. The sources of those particles were related to traffic and combustion. We had shown before (Cathomas et al. 2002) and confirm in this study that famers in our Swiss mountain area are exposed to very high concentrations of  $PM_{10}$  of another source during normal activities such as hay storage in their barns. We found that these coarse  $PM_{10}$  induced biological responses with a possible impact on health.

This  $PM_{10}$  exposure induced systemic responses such as leucocytosis. The initial increase of neutrophils with a preserved ratio of banded-to-segmented cells indicates primarily a demargination of leucocytes from the endothelium. The later relative predominance of banded neutrophils, i.e. less



**Fig. 5** Influence of  $PM_{10}$  exposure on von Willebrand factor (*above*) and coagulation factor VIII (*below*). n = 14. \* P < 0.05, \*\*\* P < 0.001 compared with baseline values (ANOVA)

mature cells, indicates a stimulation of the release from the bone marrow. In this context, the observation of increased levels of granulocyte/macrophage-colony stimulating factor (GM-CSF) after  $PM_{10}$  exposure by others is most interesting (Ishii et al. 2005).

*C*-reactive protein was increased three-fold at the last blood withdrawal, i.e. 30 h after  $PM_{10}$  exposure. CRP elevation has been described before (Ruckerl et al. 2006) with a similar time lag of 2 days, which reflects the time needed for gene transcription and de novo synthesis of CRP after triggering. The CRP increase indicates an acute phase reaction, which explains the increase of another acute phase reactant, namely fibrinogen, at the same time >24 h after  $PM_{10}$  exposure and confirms earlier observations (Ghio et al. 2000). Elevated CRP levels are associated with coronary heart disease (Danesh et al. 2000). Fibrinogen increases plasma viscosity, as seen in our study, which impairs blood flow and might be a contributing factor to the higher mortality during air pollution episodes (Peters et al. 2000).

It seems surprising that the systemic levels of inflammatory cytokines TNF-alpha and IL-6 were not affected. In vitro studies on cultured alveolar macrophages and lung epithelial cells had shown a stimulation of TNF-alpha (Jimenez et al. 2002) and IL-6 (van Eeden et al. 2001).

Platelet aggregation was increased after  $PM_{10}$  inhalation, which was seen by the faster pore closure in the PFA-100<sup>®</sup> instrument (Fig. 4) in the evening after exposure and even more marked in the next morning. Note that platelet aggregatory function is subjected to circadian changes with most activity in the morning, what we (Lehmann et al. 2006) and others (Dalby et al. 2000) had observed before. Platelet aggregation is the key event in acute occlusion of an artery. The increased aggregatory function of platelets after  $PM_{10}$ exposure may be a factor contributing to the higher incidence of cardiovascular events and mortality under these circumstances.

The PFA-100<sup>®</sup> instrument is sensitive to changes in circulating vWF, which is the bridging molecule between platelet glycoprotein Ib/IX and collagen, inducing platelet adhesion to the vessel wall. The increase in vWF observed in this study (Fig. 5) may explain the accelerated platelet aggregation in the PFA-100<sup>®</sup> instrument, and is indicative for an endothelial activation, which is in line with earlier observations (Ahmadizad et al. 2006; Törnqvist et al. 2007). Such endothelial cell activation with increase of circulating vWF is an independent risk factor for acute coronary events (Morange et al. 2004; Lee et al. 2005). The fact that sP-selectin, a marker of in vivo platelet aggregation and consumption, was hardly changed, suggests that platelet aggregation and consumption had not occurred in vivo in our healthy volunteers, despite the increase platelet aggregability, probably because they had intact vessel walls without exposure of subendothelial collagen required for platelet adhesion. The unchanged levels of TAT indicate that plasmatic coagulation was not affected, which is in general agreement with recent investigations in humans (Ruckerl et al. 2006).

The mechanisms by which  $PM_{10}$  induce these systemic reactions, which are independent of particle-induced effects on the lungs, are poorly understood. Our analysis of the particle size distribution (Fig. 1) revealed that these rural air pollutants are much larger than the particles generated by (diesel) motor vehicles and wood combustion, which measure about 10 times less. The composition of our rural  $PM_{10}$  has not been analysed so far, but may be quite different from other  $PM_{10}$ , e.g. generated by motor vehicles, and contain more organic particles. Fine and ultrafine particles may pass rapidly into the systemic circulation (Nemmar et al. 2002), although probably only as a small fraction (Mills et al. 2006). Our results would now suggest that even course particles of a few  $\mu$ m may elict systemic effects. Possible mechanisms could be a direct passage of such course PM, which, however, has not been documented so far, phagocytosis by alveolar macrophages, which then transport these particles via lymph nodes into the systemic circulation (Peters et al. 2006), or an induction of some second messenger by the PM<sub>10</sub> in alveolar epithelial cells, which would induce an indirect systemic response.

The question arises as to whether the physical activity of hay storing itself affected the systemic reactions seen in our study. It has recently been shown that moderate intensity exercise, i.e. 30 min. at 50% maximal oxygen uptake, does not influence circulating neutrophils, lymphocytes, monocytes, serum IL-6 and C-reactive protein (Markovitch et al. 2008). With regard to platelet function, moderate exercise did not affect platelet aggregation (Ahmadizad et al. 2006; Winther et al. 1992), or even decreased it, which has been shown by several groups (Petidis et al. 2008; Tozzi-Ciancarelli et al. 2002; Wang et al. 1994). Taken together, it seems rather unlikely that the moderate physical activity during hay storing contributed significantly to the measured effects, it may even have decreased the effect on platelet aggregation as suggested by the work of Petidis et al. (2008).

We conclude that acute exposure to very high concentrations of rural  $PM_{10}$  during hay storage led to systemic inflammation, increased platelet aggregation and endothelial cell activation. All these three mechanisms could contribute to cardiovascular diseases and complications. Our observations would suggest that farmers may be at risk for cardiovascular events, which has however, not been confirmed by epidemiological studies so far. A better protection by wearing a mask during work with a high  $PM_{10}$ exposure such as hay storage may thus be warranted. The traditional hay storage in the Swiss mountains resulting in very high short-term rural  $PM_{10}$ -concentrations may serve as a model to further study acute  $PM_{10}$  effects.

## References

- Ahmadizad S, El-Sayed MS, MacLaren DP (2006) Responses of platelet activation and function to a single bout of resistance exercise and recovery. Clin Hemorheol Microcirc 35:159–168
- Barnett AG, Williams GM, Schwartz J et al (2006) The effects of air pollution on hospitalizations for cardiovascular disease in elderly people in Australian and New Zealand cities. Environ Health Perspect 114:1018–1023
- Brook RD, Franklin B, Cascio W et al (2004) Air pollution and cardiovascular disease: a statement for healthcare professionals from the expert panel on population and prevention science of the American heart association. Circulation 109:2655–2671. doi:10.1161/ 01.CIR.0000128587.30041.C8

- Bünger J, Westphal G, Mönnich A et al (2004) Cytotoxicity of occupationally and environmentally relevant mycotoxins. Toxicology 202:199–211. doi:10.1016/j.tox.2004.05.007
- Cathomas RL, Bruesch H, Fehr R et al (2002) Organic dust exposure in dairy farmers in an alpine region. Swiss Med Wkly 132:174– 178
- Dalby MC, Davidson SJ, Burman JF et al (2000) Diurnal variation in platelet aggregation with the PFA-100 platelet function analyser. Platelets 11:320–324. doi:10.1080/09537100050144731
- Danesh J, Whincup P, Walker M et al (2000) Low grade inflammation and coronary heart disease: prospective study and updated metaanalyses. BMJ 321:199–204. doi:10.1136/bmj.321.7255.199
- Dockery DW, Pope CAIII, Xu X et al (1993) An association between air pollution and mortality in six U.S. cities. N Engl J Med 329:1753–1759. doi:10.1056/NEJM199312093292401
- Dominici F, McDermott A, Daniels M et al (2005) Revised analyses of the National Morbidity, Mortality, and Air Pollution Study: mortality among residents of 90 cities. J Toxicol Environ Health A 68:1071–1092. doi:10.1080/15287390590935932
- Ghio AJ, Kim C, Devlin RB (2000) Concentrated ambient air particles induce mild pulmonary inflammation in healthy human volunteers. Am J Respir Crit Care Med 162:981–988
- Gray MR, Thrasher JD, Crago R et al (2003) Mixed mold mycotoxicosis: immunological changes in humans following exposure in water-damaged buildings. Arch Environ Health 58:410–420
- Hoek G, Brunekreef B, Fischer P, van Wijnen J et al (2001) The association between air pollution and heart failure, arrhythmia, embolism, thrombosis, and other cardiovascular causes of death in a time series study. Epidemiology 12:355–357. doi:10.1097/ 00001648-200105000-00017
- Ishii H, Hayashi S, Hogg JC et al (2005) Alveolar macrophage-epithelial cell interaction following exposure to atmospheric particles induces the release of mediators involved in monocyte mobilization and recruitment. Respir Res 6:87. doi:10.1186/1465-9921-6-87
- Jimenez LA, Drost EM, Gilmour PS et al (2002) PM(10)-exposed macrophages stimulate a proinflammatory response in lung epithelial cells via TNF-alpha. Am J Physiol Lung Cell Mol Physiol 282:L237–L248
- Lanki T, Pekkanen J, Aalto P et al (2006) Associations of traffic related air pollutants with hospitalisation for first acute myocardial infarction: the HEAPSS study. Occup Environ Med 63:844–851. doi:10.1136/oem.2005.023911
- Lee KW, Blann AD, Lip GY (2005) Plasma markers of endothelial damage/dysfunction, inflammation and thrombogenesis in relation to TIMI risk stratification in acute coronary syndromes. Thromb Haemost 94:1077–1083
- Lehmann T, Mairbäurl H, Pleisch B et al (2006) Platelet count and function at high altitude and in high-altitude pulmonary edema. J Appl Physiol 100:690–694. doi:10.1152/japplphysiol.00991.2 005
- Markovitch D, Tyrrell RM, Thompson D (2008) Acute moderateintensity exercise in middle-aged men has neither an anti- nor proinflammatory effect. J Appl Physiol 105:260–265. doi:10.1152/ japplphysiol.00096.2008
- Mills NL, Amin N, Robinson SD et al (2006) Do inhaled carbon nanoparticles translocate directly into the circulation in humans? Am J Respir Crit Care Med 173:426–431. doi:10.1164/rccm.200506-865OC
- Morange PE, Simon C, Alessi MC et al (2004) Endothelial cell markers and the risk of coronary heart disease: the prospective epidemiological study of myocardial infarction (PRIME) study. Circulation 109:1343–1348. doi:10.1161/01.CIR.0000120705. 55512.EC
- Nemmar A, Hoet PH, Vanquickenborne B et al (2002) Passage of inhaled particles into the blood circulation in humans. Circulation 105:411–414. doi:10.1161/hc0402.104118

- Peters A (2005) Particulate matter and heart disease: evidence from epidemiological studies. Toxicol Appl Pharmacol 207:477–482. doi:10.1016/j.taap.2005.04.030
- Peters A, Doring A, Wichmann HE et al (2000) Increased plasma viscosity during an air pollution episode: a link to mortality? Lancet 349:1582–1587. doi:10.1016/S0140-6736(97)01211-7
- Peters A, Veronesi B, Calderon-Garciduenas L et al (2006) Translocation and potential neurological effects of fine and ultrafine particles a critical update. Part Fibre Toxicol 3:13. doi:10.1186/1743-8977-3-13
- Petidis K, Douma S, Doumas M et al (2008) The interaction of vasoactive substances during exercise modulates platelet aggregation in hypertension and coronary artery disease. BMC Cardiovasc Disord 27:8–11
- Pope CAIII, Thun MJ, Namboodiri MM et al (1995) Particulate air pollution as a predictor of mortality in a prospective study of U.S. adults. Am J Respir Crit Care Med 151:669–674
- Pope CAIII, Burnett RT, Thurston GD et al (2004) Cardiovascular mortality and long-term exposure to particulate air pollution: epidemiological evidence of general pathophysiological pathways of disease. Circulation 109:71–77. doi:10.1161/01.CIR.000010892 7.80044.7F
- Pope CAIII, Muhlestein JB, May HT et al (2006) Ischemic heart disease events triggered by short-term exposure to fine particulate air pollution. Circulation 114:2443–2448. doi:10.1161/CIRCULA-TIONAHA.106.636977
- Ruckerl R, Ibald-Mulli A, Koenig W et al (2006) Air pollution and markers of inflammation and coagulation in patients with coronary heart disease. Am J Respir Crit Care Med 173:432–441. doi:10.1164/rccm.200507-1123OC

- Salvi S, Blomberg A, Rudell B et al (1999) Acute inflammatory responses in the airways and peripheral blood after short-term exposure to diesel exhaust in healthy human volunteers. Am J Respir Crit Care Med 159:702–709
- Törnqvist H, Mills NL, Gonzalez M et al (2007) Persistent endothelial dysfunction in humans after diesel exhaust inhalation. Am J Respir Crit Care Med 176:395–400. doi:10.1164/rccm.200606-872OC
- Tozzi-Ciancarelli MG, Penco M, Di MC (2002) Influence of acute exercise on human platelet responsiveness: possible involvement of exercise-induced oxidative stress. Eur J Appl Physiol 86:266– 272. doi:10.1007/s00421-001-0542-8
- Van Eeden SF, Tan WC, Suwa T et al (2001) Cytokines involved in the systemic inflammatory response induced by exposure to particulate matter air pollutants (PM(10)). Am J Respir Crit Care Med 164:826–830
- Wang JS, Jen CJ, Kung HC (1994) Different effects of strenuous exercise and moderate exercise on platelet function in men. Circulation 90:2877–2885
- Wellenius GA, Schwartz J, Mittleman MA (2005) Air pollution and hospital admissions for ischemic and hemorrhagic stroke among medicare beneficiaries. Stroke 36:2549–2553. doi:10.1161/ 01.STR.0000189687.78760.47
- Winther K, Hillegass W, Tofler GH et al (1992) Effects on platelet aggregation and fibrinolytic activity during upright posture and exercise in healthy men. Am J Cardiol 70:1051–1055. doi:10.1016/0002-9149(92)90359-7
- Zanobetti A, Schwartz J (2005) The effect of particulate air pollution on emergency admissions for myocardial infarction: a multicity case-crossover analysis. Environ Health Perspect 113:978–982